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RECOMBINANT LIGHT CHAINS OF BOTULINUM NEUROTOXINS AND LIGHT CHAIN FUSION PROTEINS FOR USE IN RESEARCH AND CLINICAL THERAPY

This application is a divisional of U.S. patent application Ser. No. 11/293,582 filed on Dec. 2, 2005, abandoned, which is a divisional of U.S. patent application Ser. No. 10/011,588 filed Nov. 6, 2001, issued as U.S. Pat. No. 7,037,680, based on U.S. Provisional Application No. 60/246,774, filed on Nov. 6, 10 2000 and U.S. provisional Application No. 60/311,966 filed Aug. 9, 2001. U.S. patent application Ser. No. 10/011,588 is a continuation-in-part of U.S. patent application Ser. No. 09/910,186 filed Jul. 20, 2001 issued as U.S. Pat. No. 7,081, 529, which is a continuation of U.S. patent application Ser. 15 No. 09/611,419 filed Jul. 6, 2000 issued as U.S. Pat. No. 7,214,787, which is a continuation of U.S. patent application Ser. No. 08/123,975, filed Sep. 21, 1993, abandoned, wherein said application Ser. No. 09/611,419, is based on U.S. Provisional Applications Nos. 60/133,866,60/133,868, 60/133, ²⁰ 869, 60/133,865, 60/133,873, and 60/133,867, all filed May 12, 1999. All priority applications are hereby incorporated herein by reference in their entirety.

FIELD OF THE INVENTION

This invention is directed to construction, expression, and purification of synthetic DNA molecules encoding polypeptides comprising botulinum neurotoxin (BoNT) light chains. The invention is also directed to methods of vaccination 30 against botulism using the expressed peptides.

BACKGROUND OF THE INVENTION

lus Clostridium produces eight forms of antigenically distinct exotoxins. Tetanus neurotoxin (TeNT) is produced by Clostridium tetani while Clostridium botulinum produces seven different neurotoxins which are differentiated serologically by specific neutralization. The botulinum neurotoxins 40 (BoNT) have been designated as serotypes A, B, C₁, D, E, F, and G. Botulinum neurotoxins (BoNT) are the most toxic substances known and are the causative agents of the disease botulism. BoNT exert their action by inhibiting the release of the neurotransmitter acetylcholine at the neuromuscular junc- 45 tion (Habermann, E., et al., (1986), "Clostridial Neurotoxins: Handling and Action at the Cellular and Molecular Level," Cur. Top. Microbiol. Immunol., 129:93-179; Schiavo, G., et al., (1992a), "Tetanus and Botulinum-B Neurotoxins Block Neurotransmitter Release by Proteolytic Cleavage of Synap- 50 tobrevin," Nature, 359:832-835; Simpson, L. L., (1986), "Molecular Pharmacology of Botulinum Toxin and Tetanus Toxin," Annu. Rev. Pharmacol. Toxicol., 26:427-453) which leads to a state of flaccid paralysis. Indeed, only a few molecules of toxin are required to abolish the action of a nerve 55 cell. Polyclonal antibodies derived from a specific neurotoxin can neutralize the toxic effects of that toxin but will not cross-neutralize another toxin serotype. Thus, to protect against all seven toxins, one needs seven vaccines.

Human botulism poisoning is generally caused by type A, 60 B, E or rarely, by type F toxin. Type A and B are highly poisonous proteins which resist digestion by the enzymes of the gastrointestinal tract. Foodborne botulism poisoning is caused by the toxins present in contaminated food, but wound and infant botulism are caused by in vivo growth in closed wounds and the gastrointestinal tract respectively. The toxins primarily act by inhibiting the neurotransmitter acetylcholine

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at the neuromuscular junction, causing paralysis. Another means for botulism poisoning to occur is the deliberate introduction of the toxin(s) into the environment as might occur in biological warfare or a terrorist attack. When the cause of botulism is produced by toxin rather than by in vivo infection the onset of neurologic symptoms is usually abrupt and occurs within 18 to 36 hours after ingestion. The most common immediate cause of death is respiratory failure due to diaphragmatic paralysis. Home canned foods are the most common sources of toxins. The most frequently implicated toxin is toxin A, which is responsible for more than 50% of morbidity resulting from botulinum toxin.

Botulinum and tetanus neurotoxins are a new class of zincendopeptidases that act selectively at discrete sites on three synaptosomal proteins of the neuroexocytotic apparatus. See Montecucco and Schiavo, 1995, and Schiavo, 1995, for review. These neurotoxins are the most potent of all the known toxins. The botulinum neurotoxins (BoNT), designed A-G, produced by seven immunologically distinct strains of Clostridium botulinum cause death by flaccid muscle paralysis at the neuromuscular junction. Extreme toxicity of these toxins and their lability in purified preparations have limited any detailed characterizations.

These neurotoxins are expressed as 150-kDa single 25 polypeptides (termed dichains) containing a disulfide bond between the 50-kDa N-terminal light chain (LC) and the 100-kDa C-terminal heavy chain (HC). A post-translational cryptic cleavage generates the two chains connected by a disulfide bond. The LC contains the toxic, zinc-endopeptidase catalytic domain. The 100-kDa HC may be further proteolyzed into a 50-kDa N-terminal membrane-spanning domain (H_n) and a 50-kDa C-terminal receptor-binding domain (H_c).

With three functional domains, the mechanism of action of The sporulating, obligate anaerobic, gram-positive bacil- 35 these neurotoxins is multiphasic: (1) The H_c domain plays a role in binding the toxins to specific receptors located exclusively on the peripheral cholinergic nerve endings (Black and Dolly, 1986). (2) The H_n domain is believed to participate in a receptor-mediated endocytotic pore formation in an acidic environment, allowing translocation of the catalytic LC into the cytosol. Reducing the disulfide bond connecting the LC with the H upon exposure to the cytosol or within the acidic endosome (Montal et al., 1992) releases the catalytic LC into the cytosol. (3) The LC then cleaves at specific sites of one of the three different soluable NSF attachment protein receptor (SNARE) proteins, synaptobrevin, syntaxin, or synaptosomal associated protein of 25 kDa (SNAP-25) (Blasi et al., 1993; Schiavo et al., 1993, 1994; Shone et al., 1993; Foran et al., 1996). These proteins are essential for synaptic vesicle fusion in exocytosis. Their proteolysis inhibits exocytosis and blocks acetylcholine secretion, leading ultimately to muscular paralysis. The LC itself is nontoxic because it cannot translocate through the cholinergic nerve ending into the cytosol. However, in digitonin-permeabilized chromaffin cells, the LC inhibits exocytosis (Bittner et al., 1989), and direct microinjection of the LC into the cytosol results in blockage of membrane exocytosis (Bittner et al., 1989; Bi et al., 1995).

The LC of all known clostridial neurotoxins contain the sequence HExxH that is characteristic of zinc-endoproteinases (Thompson et al., 1990). The essential role of zinc on the structure and catalysis of the neurotoxins is established (Fu et al., 1998). A unique feature of the neurotoxins' protease activity is their substrate requirement. Short peptides encompassing only the cleavage sites are not hydrolyzed (Foran et al., 1994; Shone and Roberts, 1994). A specific secondary and/or tertiary structure of the substrate is most probably